

## **MIBlood-EV**

# Standardized Reporting Tool for Blood EV Research (Human)

### **STUDY INFORMATION**

<sup>1.0</sup> Manuscript title								
<sup>1.1</sup> Corresponding author (Name and Email)								
<sup>1.2</sup> Institution name								
<sup>1.3</sup> Time period of exp	eriment (years)		1	<sup>.4</sup> Number	of samples			
<sup>1.5</sup> Cargo of interest	Vesicles	Protein	RNA	DNA	Other:			
<sup>1.6</sup> Biospecimen type	Plasma	Serum	<sup>1.7</sup> Bio	specimen s	tate			
<sup>1.8</sup> Source of frozen sp	ecimens			<sup>1.9</sup> Years o	f collection	(range)		

## **BLOOD COLLECTION AND PROCESSING**

<sup>2.0</sup> Patient fasting status				<sup>2.1</sup> Fasti	ng length	(e.g. h	ours/days	5)		
<sup>2.2</sup> Anatomical access site	l access site			<sup>2.3</sup> Needle diameter (e.g				gaug	ge)	
<sup>2.4</sup> Blood volume collected	l (mL)									
<sup>2.5</sup> Plasma anticoagulant		E	EDTA	Citrate	Нера	arin	Other:			
<sup>2.6</sup> Serum tube type				<sup>2.7</sup> Seru	m clotting	g time	(minutes)			
<sup>2.8</sup> Time between collection	n and fir	st cent	trifugati	on (range	in hours)					
<sup>2.9</sup> Transport temperature			:	<sup>2.10</sup> Trans	ort condi	tion o	f tubes			
<sup>2.11</sup> Centrifuge brand and	model									
<sup>2.12</sup> Bucket rotor type				<sup>2.13</sup> Numl	per of cent	trifuga	ation cycle	s		
<sup>2.14</sup> 1 <sup>st</sup> Centrifugation speed (RCF in x g)						<sup>2.15</sup> 1 <sup>s</sup>	<sup>t</sup> Rotor br	ake		
<sup>2.16</sup> 1 <sup>st</sup> Centrifugation tem	perature			<sup>2.17</sup> <b>2</b> <sup>n</sup>	<sup>I</sup> Centrifug	gation	speed (RC	CF in	x g)	
<sup>2.18</sup> 2 <sup>nd</sup> Rotor brake			2.1	<sup>9</sup> 2 <sup>nd</sup> Cent	rifugation	n temp	erature			
<sup>2.20</sup> Additional										
processing steps										
(e.g. filtration)										
<sup>2.21</sup> Storage tubes (brand,	type, sou	irce, ca	atalog n	umber)						
<sup>2.22</sup> Storage temperature			<sup>2.23</sup> Le	ngth of s	orage (ra	nge in	years)			

### PLASMA/SERUM QUALITY CONTROL

<sup>3.0</sup> Number of freeze-thaw of	cycles (range)			
<sup>3.1</sup> Thawing temperature		<sup>3.2</sup> Thawing	duration (minutes)	

#### **Hemolysis**

<sup>3.3</sup> Presence of hemolysis			<sup>3.4</sup> Number of samples affected (e.g. <25%, 25-50%)					
<sup>3.5</sup> Method used				<sup>3.6</sup> RBC count	(Media	n, 95% Cl, N)		
<sup>3.7</sup> RBC counter bra	nd and ty	ype						
<sup>3.8</sup> Spectrophotometry hemoglobin concent				ation (g/L)				
<sup>3.9</sup> Spectrophotome	eter bran	d, mode	and					
wavelength measured (e.g. 414 nm)			m)					
<sup>3.10</sup> Hemolized sam	ples were	e discard	ded					



#### **Platelets**

<sup>3.11</sup> Presence of platelets		<sup>3.12</sup> Method used (e.g. Flow Cytometry)
<sup>3.13</sup> Marker(s) used (e.g. CD	61, CD41)	
<sup>3.14</sup> Concentration (median)	, 95% CI, N)	
<sup>3.15</sup> Platelet counter instrum	nent brand	
and type		
<sup>3.16</sup> Flow cytometer brand a	and type	
<sup>3.17</sup> Flow cytometry size and	d	
fluorescence ranges of d	letection in	
nanometers and MESF,	respectively	

### **Lipoproteins**

<sup>3.18</sup> Presence of lipoproteins	<sup>3.19</sup> Method used (WB, ELISA, FC)	
<sup>3.20</sup> Spectrophotometry L-index		
<sup>3.21</sup> Spectrophotometer brand, r	nodel and	
wavelength measured (e.g. 7	00 nm)	
<sup>3.22</sup> WB Marker(s) used (e.g. Apo	) B)	
<sup>3.23</sup> Western blot images provid	ed in manuscript?	
<sup>3.24</sup> Flow cytometry marker(s) us	sed (e.g. ApoB)	
<sup>3.25</sup> Flow cytometry concentration	on (median, 95% CI, N)	
<sup>3.26</sup> Flow cytometer brand and t	уре	
<sup>3.27</sup> Flow cytometry size and		
fluorescence ranges of detec	tion	
in nanometers and MESF,		
respectively		